

ON THE OCCURRENCE OF *TAENIOMA NANUM* (KUTZING) PAPENFUSS (RHODOPHYTA) IN PORT-CROS ISLAND (MEDITERRANEAN)

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The original locality for *Taenioma nanum* (Kützing) Papenfuss (= *T. macrourum* Schousboe ex Thuret in Bornet et Thuret) appears to have been the Gulf of Naples (KUTZING, 1849, p. 897) rather than South Africa, as pointed out by PAPANFUSS (1951); the species has been collected again near Naples by FALKENBERG (1901, p. 709, as *Taenioma macrourum*), then by LEVRING in Capri Island (1942, as *Taenioma perpusillum* J. Agardh), together with *Lophosiphonia cristata* Falkenberg, from shallow waters.

Apart from these records, *Taenioma nanum* has been detected only on very few occasions in the Mediterranean: Hrayssoun (Syria: MAYHOUB, 1976 p. 188-189, as *T. macrourum*), Cazza Island (Yugoslavia: SCHIFFNER, 1931 p. 158, as *T. macrourum*), Trstenik (Yugoslavia: SCHIFFNER, 1933 p. 293) and Lavezzi Island (Corsica: VERLAQUE and BOUDOURESQUE, 1981) (Fig. 1).

Outside the Mediterranean, *T. nanum* appears to be widespread in the warm and tropical Atlantic Ocean: Morocco, Spain, Bahamas, Venezuela, Brazil.

Taenioma nanum was collected in Port-Cros Bay (Fig. 2), in a shallow and photophilous algal turf overgrazed by two species of sea-urchins: *Arbacia lixula* (Linnaeus) and *Paracentrotus lividus* (Lamarck).

The complete list of algal species occurring in a 200 cm² sample collected at that site follows (1):

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(1) In this sample (May 1979, 0.4 m depth), every species is given a value for its cover-degree abundance: percentage of ground covered by a perpendicular projection of the aerial parts of the individual plants onto the quadrat.

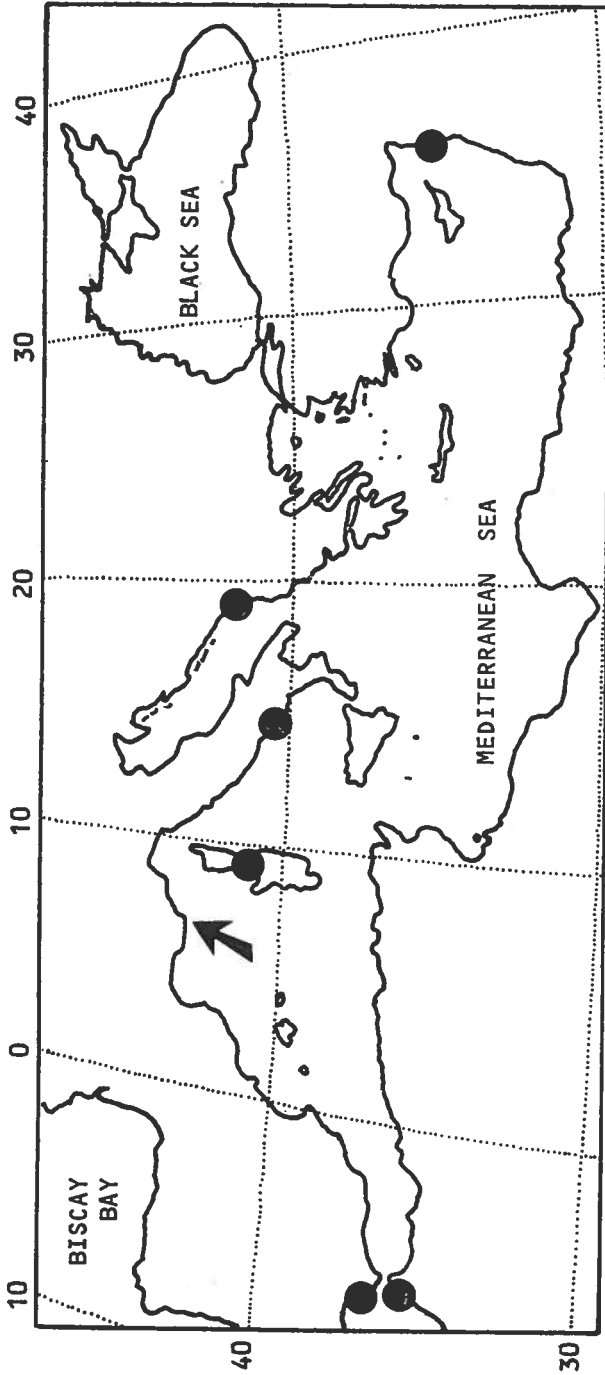


Fig. 1 : Map of Mediterranean distribution of *Taenioma nanum* (solid circles); arrow indicating the new locality. Two extra Mediterranean localities have been plotted : Tangier (Morocco) and Cadiz (Spain).

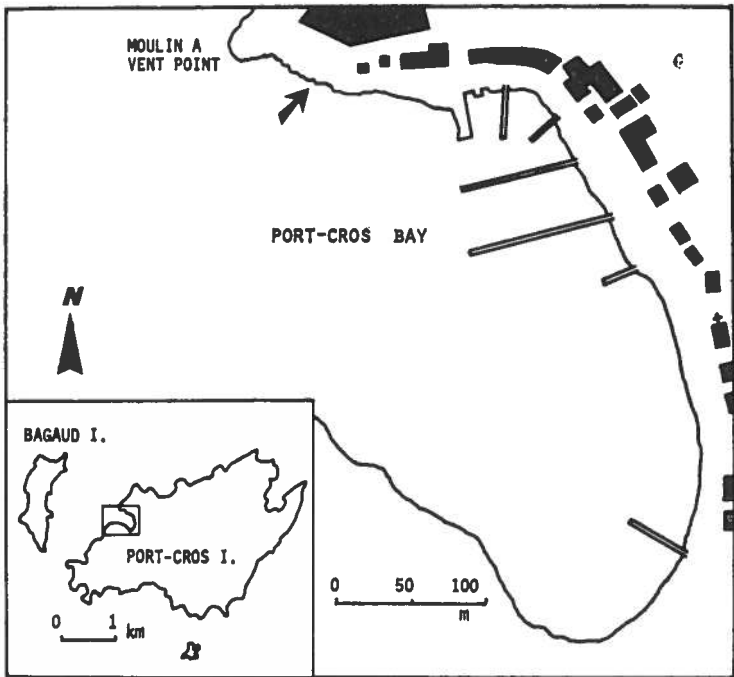


Fig. 2. Map of Port-Cros Bay showing the collecting locality (arrow).

- 40 % *Neogoniolithon notarisi* (Dufour) Setchell et Mason
- 10 % *Lithothamnium* sp.
- 10 % *Sphacelaria tribuloides* Meneghini
- 10 % *Sphacelaria cirrhosa* (Roth) C. Agardh
- 8 % *Ceramium diaphanum* (Lightfoot) Roth
- 7 % *Mesospora mediterranea* J. Feldmann
- 5 % Corallinaceae encroûtantes indéterminées
- 4 % *Corallina elongata* Ellis et Solander
- 4 % *Colpomenia sinuosa* (Mertens ex Roth) Derbès et Solier
in Castagne
- 4 % *Acinetospora vidovichii* (Meneghini) Sauvageau
- 4 % *Padina pavonica* (Linnaeus) Thivy
- 4 % *Polysiphonia tripinnata* J. Agardh
- 3 % *Dilophus fasciola* (Roth) Howe var. *repens* J. Agardh
- 2 % *Dilophus fasciola* (Roth) Howe
- 2 % *Dictyota dichotoma* (Hudson) Lamouroux
- 2 % *Phaeophila dendroides* (Crouan et Crouan) Batters
- 2 % *Fosliella zonalis* (Crouan et Crouan) J. Feldmann
- 2 % *Ceramium tenuissimum* (Lyngbye) J. Agardh

- 2 % *Enteromorpha* sp.
 1 % *Laurencia obtusa* (Hudson) Lamouroux
 1 % *Laurencia microcladia* Kützing (1)
 1 % *Acetabularia acetabulum* (Linnaeus) Silva
 1 % « *Aglaozonia parvula* (Greville) Zanardini » stadium
 1 % *Pulvinaria giraudii* (Derbès et Solier) Bourrelly
 1 % *Polysiphonia opaca* (C. Agardh) Moris et De Notaris
 0,8 % *Polysiphonia sertularoides* (Grateloup) J. Agardh
 0,7 % *Taenioma nanum* (Kützing) Papenfuss
 0,6 % *Ceramium gracillimum* Griffiths et Harvey var. *byssoideum* (Harvey) G. Mazoyer
 0,6 % *Blastophysa polymorpha* Kjellman
 0,5 % *Pseudochlorodesmis furcellata* (Zanardini) Børgesen
 0,4 % *Herposiphonia tenella* (C. Agardh) Ambronn
 0,4 % *Liagora viscida* (Forsskal) C. Agardh
 0,4 % *Cladophora* sp. 1
 0,3 % *Cladophora* sp. 2
 0,3 % *Fosliella ischiensis* Boudouresque et Cinelli
 0,3 % *Laurencia pinnatifida* (Gmelin) Lamouroux
 0,3 % *Lophosiphonia cristata* Falkenberg
 0,2 % *Erythrotrichia carnea* (Dillwyn) J. Agardh
 0,2 % *Lophosiphonia subadunca* (Kützing) Falkenberg
 0,2 % *Chondria tenuissima* (Goodenough et Woodward) C. Agardh
 0,2 % *Gelidium crinale* (Turner) Lamouroux
 0,1 % *Cystoseira balearica* Sauvageau
 0,1 % *Stictyosiphon soriferus* (Reinke) Rosenvinge
 0,1 % *Ulva lactuca* Linnaeus
 0,1 % *Fosliella farinosa* (Lamouroux) Howe
 0,1 % *Chondria dasyphylla* (Woodward) C. Agardh
 0,1 % *Boergeseniella fruticulosa* (Wulfen) Kylin
 0,1 % *Dasya ocellata* (Grateloup) Harvey in Hooker
 + *Goniotrichum alsidii* (Zanardini) Howe
 + *Acrochaetium* sp.
 + *Fosliella minutula* (Foslie) Ganesan
 + *Ceramium ciliatum* (Ellis) Ducluzeau
 + *Antithamnion cruciatum* (C. Agardh) Nägeli
 + *Dasya baillouviana* (Gmelin) Montagne

(1) Determination Marc VERLAQUE.

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